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# **Products for Biotechnology**

With Magnetic Porous Glass (MPG®)

Protocol No.: 3.206

**Product:** MPG\* Long Chain Alkylamine (30 mg/ml, 1.2 - 1.8 × 108 particles/ml)

Procedure: Covalent Attachment of Oligonucleotides.

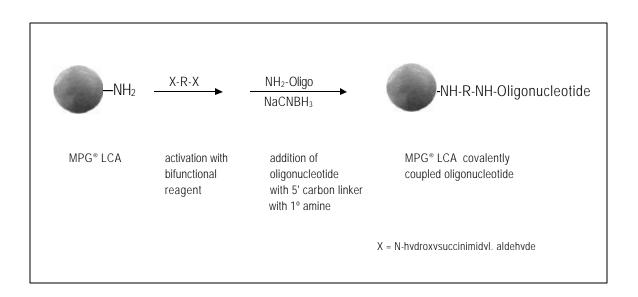
Storage: Ambient Temperature

PRODUCT

 NUMBER
 DESCRIPTION
 VOLUME

 MLCA0502
 MPG\* LCA, 5 μm, 50 nm (500 Å) pore diameter
 2 ml (60 mg)

 MLCA0510
 10 ml (300 mg)



### **General Procedure**

Materials: (Based on 10 mg MPG® Long Chain Alkylamine, suspended in 10 mM phosphate, pH 7.5, 0.15 M NaCl)

Oligonucleotide with a modified 5' end containing a 6 carbon extension ending with a primary amine

Sodium Cyanoborohydride (NaBH<sub>3</sub>CN) Sodium Phosphate, Monobasic (NaH<sub>2</sub>PO<sub>4</sub>)

Sodium Phosphate, Dibasic, Heptahydrate(Na<sub>2</sub>HPO<sub>4</sub>)

Sodium Azide (NaN<sub>3</sub>) Deionized Water (dH<sub>2</sub>O) Glycine (H<sub>2</sub>NCH<sub>2</sub>COOH) Sodium Chloride (NaCl)

25% Glutaraldehyde (CHO(CH $_2$ )  $_3$ CHO)

2N Hydrochloric Acid (HCI)

Vortex Mixer Low Speed Rotator

1.5 ml Microcentrifuge Tubes

Pipette and Pipette Tips

Magnetic Particle Separator, Prod. No. MPS0301 or

MPS0001

## Solution

Coupling Buffer (10 mM Phosphate, pH 7.5)

Activation Solution (12.5% Glutaraldehyde)

1% Sodium Cyanoborohydride Solution (Fresh)

1% Glycine Solution

Wash Buffer (10 mM Phosphate, pH 7.5, 1.0 M NaCl)

Storage Buffer (10 mM Phosphate, pH 7.5, 150 mM NaCl, 0.02% NaN<sub>3</sub>)

# **Preparation**

Dissolve 19.2 mg NaH<sub>2</sub>PO<sub>4</sub> and 225.2 mg Na<sub>2</sub>HPO<sub>4</sub>·7H<sub>2</sub>O in 80 ml dH<sub>2</sub>O. Adjust to pH 7.5 with 2N HCI, if necessary, and bring volume to 100 ml with dH<sub>2</sub>O.

Add 0.4 ml 50% Glutaraldehyde to 1.2 ml Coupling Buffer (make fresh for each reaction).

Dissolve 10 mg NaBH<sub>3</sub>CN in 1 ml Coupling Buffer.

Dissolve 10 mg Glycine in 1 ml Coupling Buffer.

Dissolve 584.7 mg NaCl in 8 ml of Coupling Buffer. Bring to 10 ml with Coupling Buffer.

Dissolve 87.7 mg NaCl, and 2 mg NaN $_3$  in 8 ml of Coupling Buffer. Bring to 10 ml with Coupling Buffer.

### Activation of MPG® Long Chain Alkylamine

- 1. Adjust the concentration of MPG® Long Chain Alkylamine to 10 mg/ml. Transfer 1 ml to a 1.5 ml microcentrifuge tube. Magnetically separate the MPG® Long Chain Alkylamine from the solution by placing the tube in a Magnetic Particle Separator for at least 30 seconds. Remove the supernatant by aspiration while the tube remains in the particle separator.
- 2. Add 1 ml of Coupling Buffer and mix well. Magnetically separate and aspirate the supernatant.
- 3. Add 250 µl of Activation Solution to the MPG\* Long Chain Alkylamine particles, mix well and place in a low speed rotator for 1½ hours at room temperature. Magnetically separate and aspirate the supernatant.
- 4. Add 1 ml of Coupling Buffer to the activated MPG® Long Chain Alkylamine particles and mix well. Magnetically separate and remove the supernatant. Repeat this step four more times.

## Coupling of Oligonucleotide to Activated MPG® Long Chain Alkylamine

- 1. Dissolve 1 mg oligonucleotide in 1 ml of Coupling Buffer. Add this mixture and 100 µl of 1% Sodium Cyanoborohydride Solution to the activated MPG\* Long Chain Alkylamine particles. Mix well and rotate 5-8 hours at room temperature. Magnetically separate and aspirate the supernatant.
- 2. Add 1 ml of 1% Glycine Solution and 100 µl of 1% Sodium Cyanoborohydride Solution, mix well and rotate 2 hours at room temperature. Magnetically separate and aspirate the supernatant.

- 3. Add 1 ml of Washing Buffer and mix well. Magnetically separate and remove the supernatant. Repeat this step five more times. The oligonucleotide-bound MPG® Long Chain Alkylamine is ready to use.
- 4. For storage, add 1 ml of Storage Buffer to the oligonucleotide-bound MPG® Long Chain Alkylamine and mix well. Magnetically separate and aspirate the supernatant. Resuspend the oligonucleotide-bound MPG® Long Chain Alkylamine particles in 1 ml Storage Buffer and store at 4°C.

This is a modification of protocol 3.1, *Covalent Attachment of Proteins*, as described by Eberwine, J. 1996 Biotechniques 20:584-591.

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